

## INTRODUCTION

Herpes simplex virus 1 and 2 (HSV-1 and HSV-2) and varicella-zoster virus (VZV) are major causes of viral meningoencephalitis [1-2]. Rapid and accurate detection of viral genomes in cerebrospinal fluid (CSF) samples is therefore mandatory for the management of patients. However, low-volume CSF may be available for molecular testing in virology laboratories.

## AIM

We evaluated the sample-to-result Simplexa™ HSV 1 & 2 Direct assay and Simplexa™ VZV Direct assay on the LIAISON® MDX platform with the use of 25 µL of sample volume, instead of 50 µL as recommended by the manufacturer DiaSorin.

## METHOD

Cycle threshold (Ct) values obtained from 25 µL and 50 µL of sample volume tested in parallel were compared for 60 HSV-positive and 60 VZV-positive different samples: 35 positive CSFs from patients, 37 negative CSFs spiked with different quantities of ATCC HSV-1, HSV-2, or VZV strains, and 48 QCMD samples. Reproducibility of Simplexa™ HSV 1 & 2 Direct assay and Simplexa™ VZV Direct assay was evaluated by intra-assay and inter-assay comparisons with the use of 25µL of sample volume.

## RESULTS

### 1- Samples tested

**Table 1. Samples used for the comparison of Ct values obtained from 25 µL and 50 µL of volume tested**

	HSV-positive samples (n=60)	VZV-positive samples (n=60)
CSF from patients	17	18
CSF spiked with ATCC viral strains <sup>a</sup>	18	19
QCMD samples <sup>b</sup>	25	23

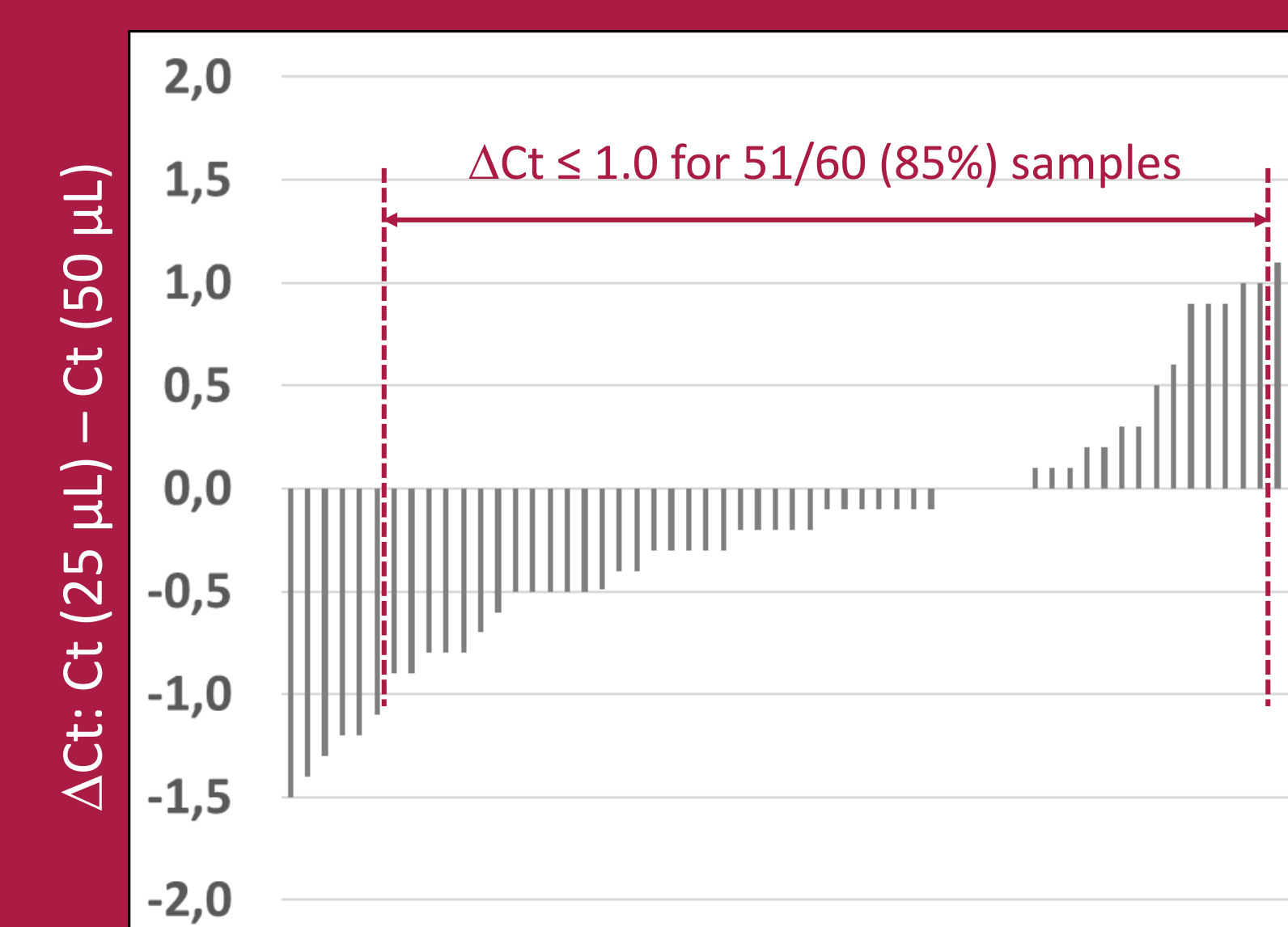
<sup>a</sup>ATCC –VR-1493: HSV-1, strain KOS; ATCC-VR-734: HSV-2, strain G; ATCC-VR-1367: VZV, strain Ellen.

<sup>b</sup>QCMD samples from Herpes simplex virus DNA EQA programmes and Varicella-Zoster virus DNA EQA, years 2018 and 2019.

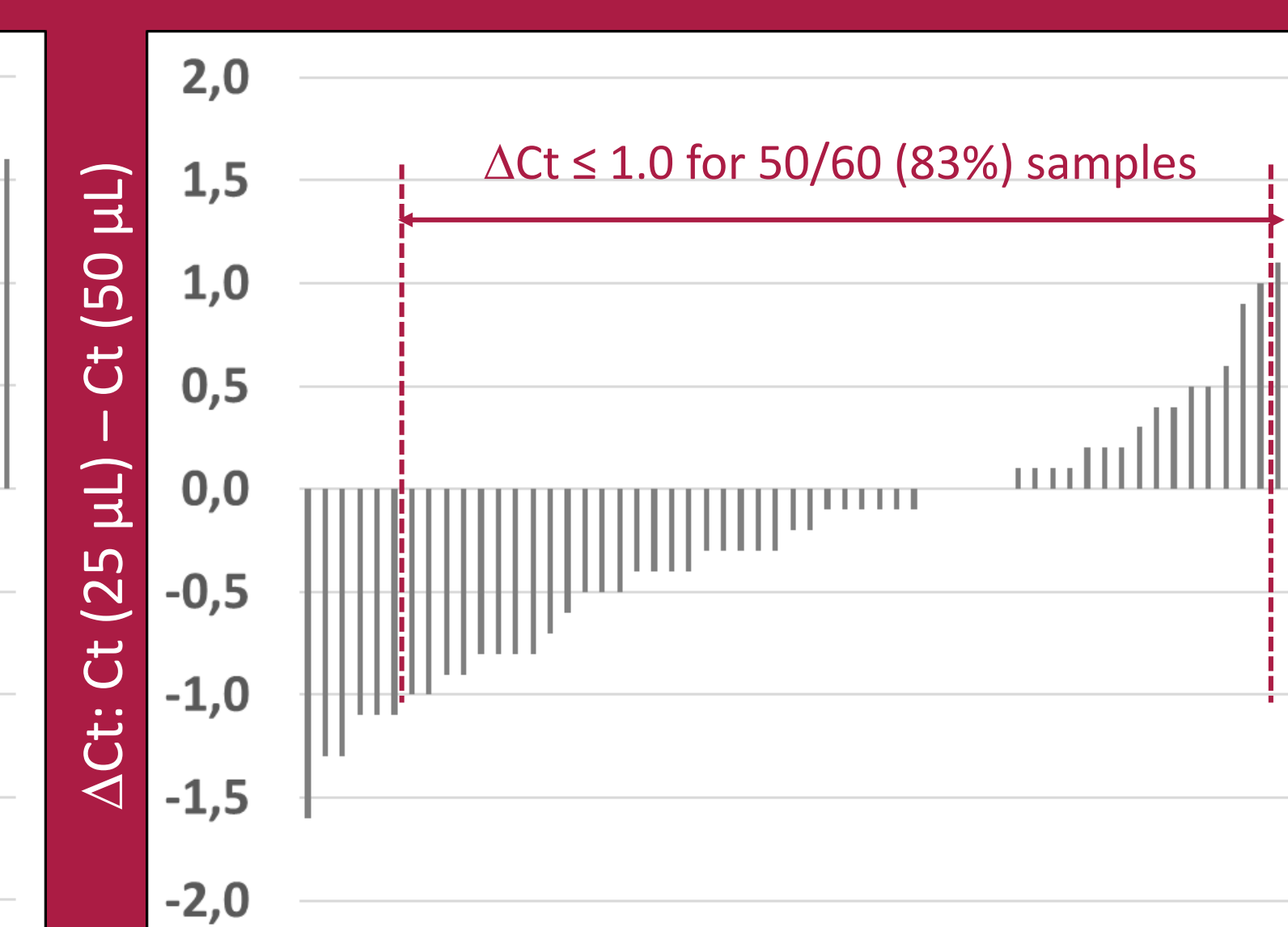
### 2- Difference of Ct values

All 120 positive samples were detected using either 25 µL or 50 µL, leading to a concordance of 100%. No PCR inhibition was observed. The difference of Ct values obtained with the 2 volumes tested ( $\Delta Ct$  [25-50]) was below or equal to 1.0 for 101 (84%) samples and ranged from 1.1 to 1.6 for the 19 (16%) remaining samples (Figure 1). Mean  $\Delta Ct$  [25-50] (SD) values were -0.1 (0.6), -0.1 (0.8), and -0.2 (0.7) for HSV-1, HSV-2, and VZV, respectively.

#### A. HSV-positive samples (n=60)



#### B. VZV-positive samples (n=60)



**Figure 1. Distribution of difference of Ct values ( $\Delta Ct$  [25-50]) for samples tested in parallel for HSV (A) and VZV (B) using either 25 µL or 50 µL of volume.**

### 3- Assay reproducibility

#### Intra-assay variability

**Method:** Simplexa™ Positive Controls for HSV-1, HSV-2, and VZV were tested 8 times with a volume of 25 µL within the same Direct Amplification Disk (DAD) during the same run on the LIAISON® MDX.

**Results:** Coefficients of variation (CVs) for intra-assay variability were 3.0%, 3.0%, and 1.9% for HSV-1, HSV-2, and VZV, respectively.

#### Inter-assay variability

**Method:** Simplexa™ Positive Controls for HSV-1, HSV-2, and VZV were tested 10 consecutive times once a week with the volume of 25 µL on the LIAISON® MDX.

**Results:** Coefficients of variation (CVs) for inter-assay variability were 1.7%, 1.0%, and 1.3% for HSV-1, HSV-2, and VZV, respectively.

## CONCLUSIONS

The results obtained in this study allow the validation of the performances of Simplexa™ HSV 1 & 2 Direct kit and Simplexa™ VZV Direct kit for the detection of HSV-1, HSV-2, and VZV genomes in CSFs with the use of 25µL of sample with no loss of sensitivity. Our results are in line with those previously obtained by Espy *et al.* [3]. This protocol will be useful for clinical laboratories to perform molecular tests with low-volume CSF samples, especially for pediatric patients.

## REFERENCES

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## CONTACT INFORMATION

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