

# QuantiFERON®-CMV assay and TTV viremia in prediction of cytomegalovirus reactivation in R+ kidney transplant recipients

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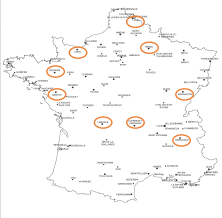
## OBJECTIVES

Monitoring CMV-specific cell-mediated immunity by the QuantiFERON®-CMV assay (QF) has been shown to be particularly useful in predicting the risk of CMV infection in kidney transplant recipients (KTR). TTV viremia has also been proposed as a biomarker of immune status in KTR. This study evaluates the ability of the QF and TTV viremia to predict CMV reactivation during the first year of transplantation in R+ KTR.

## STUDY POPULATION

A French prospective multicenter (n=9) observational study was conducted on 64 R+ KTR between 2013 and 2017.

Baseline characteristics of the study population	
Age (years; mean ± SD)	54.4 ± 13.5
Male	42 (66%)
CMV status: D+/R+ ; D-/R+	28 (44%) ; 36 (56%)
CMV prophylaxis	44 (69%)
Duration of prophylaxis (months; ± SD)	4.3 ± 1.9



## METHODS

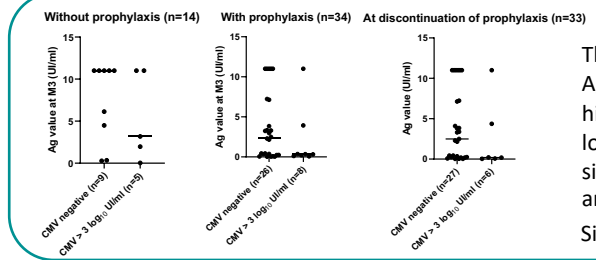
QF, TTV and CMV viral loads were performed before transplantation (J0) and from month (M) 1 to M12 after transplantation.

Quantitative values of the Ag (QF CMV T-cell specific response), Mg (QF global T-cell response) and TTV viremia were compared between CMV negative and CMV > 3 log IU/ml patients. These analyses were performed at M3 and at discontinuation of prophylaxis for each patient.

A qualitative analysis was carried out using the 0.2 IU/mL, 0.5 IU/mL and 3 log cp/ml cut-offs for Ag, Mg and TTV respectively. For markers performance evaluation, sensitivity, specificity, positive and negative predictive values (PPV and NPV) were calculated.

## RESULTS

27 patients developed CMV infection during follow-up and 15 of them exhibited a load > 3 log IU/ml. Mean TTV load peaked at M3 then decreased from M3 to M12. Both mean values of Ag and Mg decreased from J0 to M1.



## QUANTITATIVE ANALYSIS

The median of Ag at M3 as well as the median of Ag at discontinuation of prophylaxis were notably higher in CMV-negative patients than in CMV > 3 log IU/ml patients, although statistically non-significant with Fisher's test (small numbers analyzed, few CMV infections > 3 log IU/ml)

Similar results for Mg and TTV at M3 were obtained.

## QUALITATIVE ANALYSIS

**Prediction of CMV infection between M2 and M12 with a non-reactive Ag (< 0.2 IU/ml) or a non-reactive Mg (< 0.5 IU/ml) or/and a TTV viremia > 3 log cp/ml at M1 :**

	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)
<b>Individual marker analysis:</b>				
Antigen	28	81	46	67
Mitogen	30	82	50	67
TTV	67	32	36	63
<b>Combined analysis of markers:</b>				
Antigen + TTV	24	84	45	66
Mitogen + TTV	24	92	63	67

## CONCLUSION

QF and TTV at M3 or Ag value at cessation of prophylaxis do not appear to be sufficient to predict CMV reactivation and to adapt antiviral prophylaxis in R+ KTR. They are more predictive of CMV viremia control than reactivation (NPV > NPP). TTV viremia at M1 seems to be a relevant indicator of immunosuppression in patients with CMV reactivation (sensitivity of 67%). QF in combination with TTV viremia at M1 provides greater specificities and may be useful to identify patients at lower risk of CMV reactivation.